09/763,324

(FILE 'HOME' ENTERED AT 16:00:18 ON 19 MAY 2002)

FILE 'CAPLUS, BIOSIS, MEDLINE' ENTERED AT 16:00:48 ON 19 MAY 2002
L1 8 S CHITOSAN? AND HOMOGENE? AND (ENZYME? OR TYROSINASE OR PHENOL(
L2 5 DUP REM L1 (3 DUPLICATES REMOVED)

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FILE 'BIOSIS' ENTERED AT 16:00:48 ON 19 MAY 2002 COPYRIGHT (C) 2002 BIOLOGICAL ABSTRACTS INC. (R)

FILE 'MEDLINE' ENTERED AT 16:00:48 ON 19 MAY 2002

=> s chitosan? and homogene? and (enzyme? or tyrosinase or phenol(3a)oxidase? or polyphenol(3a)oxidase?) and phenol?

OR IS NOT A RECOGNIZED COMMAND

The previous command name entered was not recognized by the system. For a list of commands available to you in the current file, enter "HELP COMMANDS" at an arrow prompt (=>).

=> s chitosan? and homogene? and (enzyme? or tyrosinase or phenol(3a)oxidase? or polyphenol(3a)oxidase?) and phenol?

8 CHITOSAN? AND HOMOGENE? AND (ENZYME? OR TYROSINASE OR PHENOL(3A) OXIDASE? OR POLYPHENOL(3A) OXIDASE?) AND PHENOL?

=> dup rem l1
PROCESSING COMPLETED FOR L1

L2 5 DUP REM L1 (3 DUPLICATES REMOVED)

=> d l2 abs ibib kwic 1-5

L2 ANSWER 1 OF 5 CAPLUS COPYRIGHT 2002 ACS

A variety of crosslinked polymers exist in nature and crosslinking confers AΒ important functional properties to biomaterials. In nature, crosslinking is generally achieved by the enzymic conversion of phenols into reactive intermediates (e.g. quinones or free radicals) that undergo subsequent non-enzymic reactions. Although enzymic crosslinking reactions offer interesting opportunities for conferring functionality, it has been difficult to control the poorly characterized non-enzymic reactions. report the use of combinatorial approaches to learn how reactions of enzymically-generated quinones can be exploited for polysaccharide crosslinking and for the coupling of proteins to polysaccharides. Specifically we used tyrosinase to convert natural phenols into reactive quinones and examd. the crosslinking of the amino-polysaccharide chitosan and the coupling of chitosan to various proteins. In studies on polymer crosslinking we performed homogeneous reactions and examd. various phenols and reaction conditions. Screening was based on a rapid
method to characterize the mech. properties of the enzymically crosslinked chitosan gels. For chitosan-protein coupling we used heterogeneous conditions and screened various phenols and reaction conditions to identify conditions that coupled protein to chitosan films while maintaining biol. activity.

ACCESSION NUMBER:

2001:197377 CAPLUS

TITLE:

Combinatorial approach to biopolymer coupling and

crosslinking

AUTHOR (S):

Payne, Gregory F.; Chen, Tianhong; Vazquez-Duhalt,

Rafael; Bentley, William E.; Smith, Paul J.

CORPORATE SOURCE:

Center for Agricultural Biotechnology, University of

Maryland, College Park, MD, 20742-4450, USA

09/763,324

SOURCE:

Abstr. Pap. - Am. Chem. Soc. ((2001), 221st, BIOT-070

CODEN: ACSRAL; ISSN: 0065-7727

PUBLISHER: DOCUMENT TYPE: American Chemical Society Journal; Meeting Abstract

English

LANGUAGE:

A variety of crosslinked polymers exist in nature and crosslinking confers important functional properties to biomaterials. In nature, crosslinking is generally achieved by the enzymic conversion of phenols into reactive intermediates (e.g. quinones or free radicals) that undergo subsequent non-enzymic reactions. Although enzymic crosslinking reactions offer interesting opportunities for conferring functionality, it has been difficult to control the poorly characterized non-enzymic reactions. report the use of combinatorial approaches to learn how reactions of enzymically-generated quinones can be exploited for polysaccharide crosslinking and for the coupling of proteins to polysaccharides. Specifically we used tyrosinase to convert natural phenols into reactive quinones and examd. the crosslinking of the amino-polysaccharide chitosan and the coupling of chitosan to various proteins. In studies on polymer crosslinking we performed homogeneous reactions and examd. various phenols and reaction conditions. Screening was based on a rapid method to characterize the mech. properties of the enzymically crosslinked chitosan gels. For chitosan-protein coupling we used heterogeneous conditions and screened various phenols and reaction conditions to identify conditions that coupled protein to chitosan films while maintaining biol. activity.

L2 ANSWER 2 OF 5 CAPLUS COPYRIGHT 2002 ACS

AB A homogeneous-phase enzyme-catalyzed process for producing modified chitosan polymers or oligomers comprises reacting an enzyme, e.g., tyrosinase, with a phenolic substrate, e.g., chlorogenic acid, in the presence of a chitosan polymer or oligomer. The modified chitosan polymers or oligomers produced by the novel processes, in particular those having useful functional properties, e.g., base soly. and/or high viscosity are also claimed.

ACCESSION NUMBER:

2000:144914 CAPLUS

DOCUMENT NUMBER:

132:182264

TITLE:

Modified chitosan polymers and enzymic

methods for their production Kumar, Guneet; Payne, Gregory F.

INVENTOR(S):
PATENT ASSIGNEE(S):

USA

SOURCE:

PCT Int. Appl., 47 pp.

CODEN: PIXXD2

DOCUMENT TYPE:

Patent

LANGUAGE:

English

FAMILY ACC. NUM. COUNT:

PATENT INFORMATION:

PATENT NO. KIND DATE

APPLICATION NO. DATE

WO 2000011038 A1 20000302 WO 1999-US19106 19990820

W: AE, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CU, CZ, DE, DK, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, UA, UG, US, UZ, VN, YU, ZA, ZW, AM, AZ, BY, KG, KZ, MD, RU, TJ, TM

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RW: GH, GM, KE, LS, MW, SD, SL, SZ, UG, ZW, AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE, BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG
     AU 9957814
                        A1
                             20000314
                                            AU 1999-57814
                                                               19990820
     EP 1137673
                        A1
                             20011004
                                             EP 1999-945134
                                                               19990820
         R: AT, BE, CH, DE, DK, ES, FR, GB, GR, IT, LI, LU, NL, SE, MC, PT,
             IE, SI, LT, LV, FI, RO
PRIORITY APPLN. INFO.:
                                          US 1998-97709P
                                                           P 19980821
                                          WO 1999-US19106 W 19990820
REFERENCE COUNT:
                                THERE ARE 4 CITED REFERENCES AVAILABLE FOR THIS
                                RECORD. ALL CITATIONS AVAILABLE IN THE RE FORMAT
TI
     Modified chitosan polymers and enzymic methods for their
     production
AΒ
     A homogeneous-phase enzyme-catalyzed process for
     producing modified chitosan polymers or oligomers comprises
     reacting an enzyme, e.g., tyrosinase, with a
     phenolic substrate, e.g., chlorogenic acid, in the presence of a
     chitosan polymer or oligomer. The modified chitosan
     polymers or oligomers produced by the novel processes, in particular those
     having useful functional properties, e.g., base soly. and/or high
     viscosity are also claimed.
ST
     chitosan reaction oxidized phenol enzyme
     oxidant; tyrosinase oxygen oxidn chlorogenic acid
     chitosan modification
ΙT
     Oxidation
        (enzymic, of phenols; enzymic methods for the manuf. of
        chitosan polymers modified with oxidized phenols)
IT
     Phenols, preparation
     RL: IMF (Industrial manufacture); PREP (Preparation)
        (reaction products; enzymic methods for the manuf. of chitosan
        polymers modified with oxidized phenols)
IT
     9002-10-2, Tyrosinase
     RL: BPR (Biological process); BSU (Biological study, unclassified); BIOL
     (Biological study); PROC (Process)
        (enzymic methods for the manuf. of chitosan polymers modified
        with oxidized phenols)
İΤ
     51-61-6DP, Dopamine, oxidized, reaction products with chitosan
     106-44-5DP, p-Cresol, oxidized, reaction products with chitosan
     120-80-9DP, Catechol, oxidized, reaction products with chitosan
     327-97-9DP, Chlorogenic acid, oxidized, reaction products with
     chitosan
               9012-76-4DP, Chitosan, reaction products with
     oxidized phenols
     RL: IMF (Industrial manufacture); PREP (Preparation)
        (enzymic methods for the manuf. of chitosan polymers modified
        with oxidized phenols)
     7782-44-7, Oxygen, uses
IT
     RL: NUU (Other use, unclassified); USES (Uses)
        (enzymic methods for the manuf. of chitosan polymers modified .
        with oxidized phenols)
L2
     ANSWER 3 OF 5 CAPLUS COPYRIGHT 2002 ACS
                                                         DUPLICATE 1
     An enzymic method to graft hexyloxyphenol onto the biopolymer
AB
     chitosan was studied. The method employs tyrosinase to
     convert the phenol into a reactive o-quinone, which undergoes
     subsequent nonenzymic reaction with chitosan. Reactions were
     conducted under heterogeneous conditions using chitosan films
     and also under homogeneous conditions using aq. methanolic
```

mixts. capable of dissolving both hexyloxyphenol and chitosan.

Tyrosinase was shown to catalyze the oxidn. of hexyloxyphenol in such aq. methanolic solns. Chem. evidence for covalent grafting onto chitosan was provided by three independent spectroscopic approaches. Specifically, enzymic modification resulted in (1) the appearance of broad absorbance in the 350-nm region of the UV/vis spectra for chitosan films; (2) changes in the NH bending and stretching regions of chitosan's IR spectra; and (3) a base-sol. material with 1H-NMR signals characteristic of both chitosan and the alkyl groups of hexyloxyphenol. Hexyloxyphenol modification resulted in dramatic changes in chitosan's functional properties. On the basis of contact angle measurements, heterogeneous modification of a chitosan film yielded a hydrophobic surface.

Homogeneously modified chitosan offered rheol.

properties characteristic of assocq. water-sol. polymers.

ACCESSION NUMBER: 2000:816746 CAPLUS

DOCUMENT NUMBER: 134:99633

TITLE: Enzymatic grafting of hexyloxyphenol onto

chitosan to alter surface and rheological

properties

Chen, Tianhong; Kumar, Guneet; Harris, Michael T.; AUTHOR (S):

Smith, Paul J.; Payne, Gregory F.

CORPORATE SOURCE: Center for Agricultural Biotechnology, University of

Maryland, College Park, MD, 20742, USA

SOURCE: Biotechnology and Bioengineering (2000), 70(5),

564-573

CODEN: BIBIAU; ISSN: 0006-3592

PUBLISHER: John Wiley & Sons, Inc.

DOCUMENT TYPE: Journal

LANGUAGE: English

REFERENCE COUNT: 56 THERE ARE 56 CITED REFERENCES AVAILABLE FOR THIS

RECORD. ALL CITATIONS AVAILABLE IN THE RE FORMAT

Enzymatic grafting of hexyloxyphenol onto chitosan to alter ΤI surface and rheological properties

AΒ An enzymic method to graft hexyloxyphenol onto the biopolymer chitosan was studied. The method employs tyrosinase to convert the phenol into a reactive o-quinone, which undergoes subsequent nonenzymic reaction with chitosan. Reactions were conducted under heterogeneous conditions using chitosan films and also under homogeneous conditions using aq. methanolic mixts. capable of dissolving both hexyloxyphenol and chitosan. Tyrosinase was shown to catalyze the oxidn. of hexyloxyphenol in such aq. methanolic solns. Chem. evidence for covalent grafting onto chitosan was provided by three independent spectroscopic approaches. Specifically, enzymic modification resulted in (1) the appearance of broad absorbance in the 350-nm region of the UV/vis spectra for chitosan films; (2) changes in the NH bending and stretching regions of chitosan's IR spectra; and (3) a base-sol. material with 1H-NMR signals characteristic of both chitosan and the alkyl groups of hexyloxyphenol. Hexyloxyphenol modification resulted in dramatic changes in chitosan's functional properties. On the basis of contact angle measurements, heterogeneous modification of a chitosan film yielded a hydrophobic surface. Homogeneously modified chitosan offered rheol.

properties characteristic of assocg. water-sol. polymers.

ST tyrosinase grafting hexyloxyphenol chitosan

ΙT Contact angle Viscosity

(enzymic grafting of hexyloxyphenol onto chitosan to alter

AUTHOR(S):

```
surface and rheol. properties)
IT
     Oxidation
        (enzymic; enzymic grafting of hexyloxyphenol onto chitosan to
        alter surface and rheol. properties)
     Polymers, preparation
     RL: BMF (Bioindustrial manufacture); BPN (Biosynthetic preparation); PRP
     (Properties); PUR (Purification or recovery); BIOL (Biological study);
     PREP (Preparation)
        (graft; enzymic grafting of hexyloxyphenol onto chitosan to
        alter surface and rheol. properties)
IT
     9012-76-4DP, Chitosan, graft copolymer with hexyloxyphenol
     RL: BMF (Bioindustrial manufacture); BPN (Biosynthetic preparation); PRP
     (Properties); PUR (Purification or recovery); BIOL (Biological study);
     PREP (Preparation)
        (enzymic grafting of hexyloxyphenol onto chitosan to alter
        surface and rheol. properties)
IT.
     9002-10-2, Tyrosinase
     RL: BPR (Biological process); BSU (Biological study, unclassified); CAT
     (Catalyst use); BIOL (Biological study); PROC (Process); USES (Uses)
        (enzymic grafting of hexyloxyphenol onto chitosan to alter
        surface and rheol. properties)
IT
     622-62-8
               9012-76-4, Chitosan 18979-55-0, 4-n-Hexyloxyphenol
     26638-03-9, Methoxyphenol
     RL: BPR (Biological process); BSU (Biological study, unclassified); RCT
     (Reactant); BIOL (Biological study); PROC (Process); RACT (Reactant or
        (enzymic grafting of hexyloxyphenol onto chitosan to alter
        surface and rheol. properties)
IT
     320401-59-0
     RL: BSU (Biological study, unclassified); MFM (Metabolic formation); BIOL
     (Biological study); FORM (Formation, nonpreparative)
        (enzymic grafting of hexyloxyphenol onto chitosan to alter
        surface and rheol. properties)
IT
     69818-23-1
                  320401-57-8
     RL: BSU (Biological study, unclassified); MFM (Metabolic formation); RCT
     (Reactant); BIOL (Biological study); FORM (Formation, nonpreparative);
     RACT (Reactant or reagent)
        (enzymic grafting of hexyloxyphenol onto chitosan to alter
        surface and rheol. properties)
     ANSWER 4 OF 5 CAPLUS COPYRIGHT 2002 ACS
L2
     It was obsd. that addn. of tyrosinase and the simple
     phenol, p-cresol, to semi-dil. solns. of chitosan (I)
     resulted in the in situ formation of I gels. Specifically,
     homogeneous reactions were conducted with I solns. (0.32 w/v %) at
     pH near 6.0 and with cresol levels of 0.6 molar equiv (relative to I amino
     groups). Oscillatory shear measurements showed that the enzymic reaction
     resulted in large increases in the complex viscosity (.eta.*) and storage
     and loss moduli (G' and G''). These dynamic measurements indicated that
     the enzymic reaction resulted in the conversion of the nearly Newtonian
     semi-dil. I solns. into gels. The rheol. behavior of these
     enzymically-generated gels was compared to the behavior of acidic I solns.
     and to solns. contg. xanthan gum.
ACCESSION NUMBER:
                         2000:450035 CAPLUS
DOCUMENT NUMBER:
                         134:6082
TITLE:
                         In situ chitosan gelation using the
                         enzyme tyrosinase
```

Kumar, G.; Bristow, J. F.; Smith, P. J.; Payne, G. F.

09/763,324

CORPORATE SOURCE: Center for Agricultural Biotechnology, Univ. Maryland,

College Park, MD, 20742, USA

SOURCE: Advances in Chitin Science (2000), 4 (EUCHIS'99),

345-348

CODEN: ACSCFF

PUBLISHER: Universitaet Potsdam, Universitaetsbibliothek

DOCUMENT TYPE: Journal LANGUAGE: English

REFERENCE COUNT: 18 THERE ARE 18 CITED REFERENCES AVAILABLE FOR THIS

RECORD. ALL CITATIONS AVAILABLE IN THE RE FORMAT

TI In situ chitosan gelation using the enzyme tyrosinase

AB It was obsd. that addn. of tyrosinase and the simple phenol, p-cresol, to semi-dil. solns. of chitosan (I) resulted in the in situ formation of I gels. Specifically, homogeneous reactions were conducted with I solns. (0.32 w/v %) at pH near 6.0 and with cresol levels of 0.6 molar equiv (relative to I amino groups). Oscillatory shear measurements showed that the enzymic reaction resulted in large increases in the complex viscosity (.eta.\*) and storage and loss moduli (G' and G''). These dynamic measurements indicated that the enzymic reaction resulted in the conversion of the nearly Newtonian semi-dil. I solns. into gels. The rheol. behavior of these enzymically-generated gels was compared to the behavior of acidic I solns. and to solns. contg. xanthan gum.

ST tyrosinase enzyme cresol in situ gelation chitosan soln; mech loss viscoelasticity viscosity chitosan soln gelation cresol enzyme

IT Gelation

Mechanical loss Viscoelasticity

Viscosity

(in situ chitosan soln. gelation using tyrosinase enzyme and p-cresol)

IT Enzymes, uses

RL: NUU (Other use, unclassified); USES (Uses) (tyrosinase; in situ chitosan soln. gelation using tyrosinase enzyme and p-cresol)

IT 106-44-5, p-Cresol, uses 9002-10-2, Tyrosinase
RL: NUU (Other use, unclassified); USES (Uses)
 (in situ chitosan soln. gelation using tyrosinase
 enzyme and p-cresol)

IT 9012-76-4, Chitosan

RL: PEP (Physical, engineering or chemical process); PRP (Properties); PROC (Process)

(in situ chitosan soln. gelation using tyrosinase enzyme and p-cresol)

L2 ANSWER 5 OF 5 CAPLUS COPYRIGHT 2002 ACS DUPLICATE 2

AB Chitosan (I) is a natural biopolymer whose rich amine functionality confers water soly. at low pH. At higher pH's (>6.5), the amines are deprotonated, and I is insol. To attain water soly. under basic conditions the hydrophilic compd. chlorogenic acid (II) was enzymically grafted onto I. Despite its name, II is a non-chlorinated phenolic natural product that has carboxylic acid and OH functionality. The enzyme used was tyrosinase, which converts a wide range of phenolic substrates into electrophilic o-quinones. The o-quinones are freely diffusible and can undergo reaction with the nucleophilic amino groups of I. Using slightly acidic conditions

ΤI

AB

(pH = 6.0), it was possible to modify I under homogeneous conditions. When the amt. of II used in the modification reaction was >30% relative to the I amino groups, the modified I was obsd. to be sol. under both acidic and basic conditions, and to have a pH window of insoly. at near neutral pH. Proton NMR spectra confirmed that I was chem. modified, although the degree of modification was low. ACCESSION NUMBER: 1999:131727 CAPLUS DOCUMENT NUMBER: 130:239087 TITLE: Enzymic grafting of a natural product onto chitosan to confer water solubility under basic conditions AUTHOR(S): Kumar, Guneet; Smith, Paul J.; Payne, Gregory F. Center for Agricultural Biotechnology, University of CORPORATE SOURCE: Maryland, College Park, MD, 20742, USA Biotechnology and Bioengineering (1999), 63(2), SOURCE: 154-165 CODEN: BIBIAU; ISSN: 0006-3592 PUBLISHER: John Wiley & Sons, Inc. DOCUMENT TYPE: Journal LANGUAGE: English REFERENCE COUNT: THERE ARE 56 CITED REFERENCES AVAILABLE FOR THIS 56 RECORD. ALL CITATIONS AVAILABLE IN THE RE FORMAT Enzymic grafting of a natural product onto chitosan to confer water solubility under basic conditions Chitosan (I) is a natural biopolymer whose rich amine functionality confers water soly. at low pH. At higher pH's (>6.5), the amines are deprotonated, and I is insol. To attain water soly. under basic conditions the hydrophilic compd. chlorogenic acid (II) was enzymically grafted onto I. Despite its name, II is a non-chlorinated phenolic natural product that has carboxylic acid and OH functionality. The enzyme used was tyrosinase, which converts a wide range of phenolic substrates into electrophilic o-quinones. The o-quinones are freely diffusible and can undergo reaction with the nucleophilic amino groups of I. Using slightly acidic conditions (pH = 6.0), it was possible to modify I under homogeneous conditions. When the amt. of II used in the modification reaction was >30% relative to the I amino groups, the modified I was obsd. to be sol. under both acidic and basic conditions, and to have a pH window of insoly. at near neutral pH. Proton NMR spectra confirmed that I was chem. modified, although the degree of modification was low.

ST chlorogenic acid grafting chitosan soly tyrosinase enzyme catalyst

IT Solubility

> (alk.; enzymically catalyzed grafting of natural products onto chitosan to confer water soly. under alk. conditions)

ΙT Enzymes, uses

RL: CAT (Catalyst use); USES (Uses)

(enzymically catalyzed grafting of natural products onto chitosan to confer water soly. under alk. conditions)

IT Polymerization

Polymerization catalysts

(graft; enzymically catalyzed grafting of natural products onto chitosan to confer water soly. under alk. conditions)

TΤ 9002-10-2, Tyrosinase

RL: CAT (Catalyst use); USES (Uses)

(enzymically catalyzed grafting of natural products onto chitosan to confer water soly. under alk. conditions)

IT 327-97-9, Chlorogenic acid 9012-76-4, Chitosan RL: RCT (Reactant); RACT (Reactant or reagent) (enzymically catalyzed grafting of natural products onto chitosan to confer water soly. under alk. conditions)

L Number	Hits	Search Text	DB	Time stamp
1	12	chitosan\$2 same (polymer\$2 or copolymer\$2) same viscosity	USPAT;	2002/05/19 15:46
		same (cps! or centipoise\$2 or poise\$2)	US-PGPUB	